

Estimation of hemolymph a_{CO_2} and pKapp in disk abalone *Haliotis (Nordotis) discus discus* between 10°C and 20°C

Takeshi Handa[†] and Akira Araki

Abstract: We investigated the effect of temperature on disk abalone *Haliotis (Nordotis) discus discus* hemolymph CO_2 solubility coefficient (a_{CO_2}) and the apparent dissociation constant of carbonic acid (pKapp). The disk abalone hemolymph was equilibrated with a standard CO_2 gas mixture between 10°C and 20°C, to obtain expressions for a_{CO_2} and pKapp as a function of temperature. The relationship between a_{CO_2} and temperature (T) is expressed as follows: $a_{CO_2} = 74.005 - 1.2936 \cdot T - 0.00944 \cdot T^2$. And the relationship between pKapp and temperature expressed as follows: $pKapp = 6.4675 - 0.08682 \cdot T + 0.003996 \cdot T^2$. In these equations, the parameter units are °C for T and $\mu M/L/torr$ for a_{CO_2} . The non-bicarbonate buffer values (β_{NB}), obtained as a regression coefficient relating pH and $[HCO_3^-]$, were 2.5 Slykes at 10°C, 2.2 Slykes at 15°C, and 3.4 Slykes at 20°C. These equations enable estimation of hemolymph a_{CO_2} and pKapp between temperatures of 10°C and 20°C.

Key words: *Haliotis (Nordotis) discus discus*, disk abalone, hemolymph acid–base balance, CO_2 solubility (a_{CO_2}), apparent dissociation constant (pKapp), temperature effects

Introduction

Disk abalone *Haliotis (Nordotis) discus discus* is a marine mollusc classified in the Haliotidae, Vetigastropoda, GASTROPODA, and inhabits intertidal zones at a depth of about 20 m throughout the whole of the Japan Sea and the Pacific coast from Ibaraki Prefecture to Kyushu¹⁾. In Japan, the disk abalone is traded as an economical importance species, and was previously evaluated in terms of juvenile abalone growth²⁻⁴⁾, ammonia excretion⁵⁾, oxygen consumption⁵⁾, amyotrophia⁶⁻⁹⁾, and immune responses to bacterial and viral stresses^{10,11)}. The anatomical structures of the digestive diverticula, ctenidium, and circulatory system are also well studied in this species^{12,13)}. Gill ventilation volume regulation and O_2 uptake in the disk abalone was evaluated for both normoxic and hypoxic conditions¹⁴⁻¹⁷⁾. The disk abalone hemolymph acid–base balance under resting conditions was also studied¹⁸⁾. In normoxic and normocapnic seawater at 28°C, the disk abalone has a hemolymph pH

of 7.320 and total CO_2 concentration (T_{CO_2}) of 1.78 mM/L¹⁸⁾. The CO_2 partial pressure (P_{CO_2}) and bicarbonate concentration ($[HCO_3^-]$) for the hemolymph, calculated by the Henderson–Hasselbalch equation, was reported as 4.2 torr and 1.63 mM/L, respectively¹⁸⁾. The Henderson–Hasselbalch equation is often used in studies involving acid–base balance to calculate P_{CO_2} owing to the relative ease and accuracy of this method¹⁹⁾. In the equation, the CO_2 solubility coefficient (a_{CO_2}) and apparent dissociation constant of carbonic acid (pKapp) of the hemolymph are required for the experimental animals. The a_{CO_2} and pKapp vary with temperature¹⁹⁾ but there are few reports on the effect of temperature on the hemolymph a_{CO_2} and pKapp in Haliotidae. Therefore, we undertook a preliminary investigation to evaluate the effect of temperature on disk abalone hemolymph a_{CO_2} and pKapp. If the relationships between temperature on the a_{CO_2} and pKapp are simple for disk abalone hemolymph, the P_{CO_2} and $[HCO_3^-]$ may be easily calculated, which is useful in understanding the hemolymph acid–base balance,

Received date: 29 November 2023, Accepted date 16 January 2024

Affiliation: Department of Applied Aquabiology, National Fisheries University, Nagata-honmachi, Shimonoseki, Yamaguchi Pref., JAPAN

[†] Corresponding author: handat@fish-u.ac.jp (T. HANDA)

respiratory physiology, and interpreting the effects of temperature on disk abalone aquaculture environments. In this study, we equilibrated disk abalone hemolymph with CO₂ standard gas mixes between 10°C and 20°C to obtain relational expressions between the coefficients (*aco*₂ and pK_{app}) and temperature.

Materials and Methods

Experimental animals and conditions

These experiments used 102 disk abalone *Haliotis (Nardotis) discus discus* (total wet weight: 97.8 ± 16.6 g (mean ± SD)). The animals were obtained from a commercial marine farm in Yamaguchi prefecture, Japan. After cleaning the shell surface, the animals were reared over 1 month in aerated seawater at 10°C, 15°C, or 20°C and fed seaweed (*Sargassum macrocarpum*, *Ecklonia kurome* and *Ulva pertusa*). Twenty-four hours before hemolymph collection, the disk abalone were transferred to particle-free (>0.45 μm) seawater without seaweed. All experiments were conducted in seawater with a salinity of 30 psu, O₂ saturation 98%, pH 8.0, and total CO₂ concentration 1.5 mM/L.

Animal surgery and hemolymph collection

Disk abalone, reared at each temperature (10°C, 15°C, or 20°C), were subjected to a surgical operation to collect the hemolymph¹⁸⁾. The disk abalone was submerged in MgCl₂ solution (29-31 psu) to prevent muscle contraction²⁰⁾. After muscle relaxation, a polyethylene tube (0.96 mm outer diameter, 0.58 mm inner diameter, PE-50, Becton Dickinson CO.) was inserted into the vein located near the margin of the shell¹⁸⁾. The cannulated animals were then transferred to a respiratory chamber supplied with flowing aerated water, and allowed to recover overnight at the assigned temperatures. Hemolymph was collected through the cannula for *in vitro* experiments.

Experimental protocols

Hemolymph *aco*₂ was determined by first adjusting to

pH 7.5 by the addition of lactic acid (Wako Pure Chemical Industries, Ltd.). The hemolymph with lactic acid was centrifuged, and the supernatant used for *aco*₂ analysis. The supernatant sample was transferred to a tonometer flask and equilibrated with humidified standard CO₂ gas (CO₂, 5.0% or 15.0%; O₂, 20.9%; N₂ Balance) using an equilibrator (DEQ-1, Cameron Instruments) at 10°C, 15°C, or 20°C, and measuring the Tco₂ for each equilibrated sample. The Pco₂ of the equilibrated sample was calculated using the known standard gas CO₂ concentration, barometric pressure, and water vapor pressure at each experimental temperature. The *aco*₂ was calculated using the equation:

$$aco_2 = Tco_2 \cdot Pco_2^{-1}$$

For pK_{app} determination, the hemolymph was transferred to a tonometer flask and equilibrated with humidified standard CO₂ gases (CO₂, 0.1-5.0%; O₂, 20.9%; N₂ Balance) using an equilibrator at 10°C, 15°C, or 20°C. After equilibration, the pH and Tco₂ of the sample were measured. Using the sample pH, Tco₂, and *aco*₂ which calculated from the above equation, pK_{app} was determined by rearrangement of the Henderson-Hasselbalch equation^{19,21)} as follows:

$$pK_{app} = pH - \log [(Tco_2 - aco_2 \cdot Pco_2) \cdot (aco_2 \cdot Pco_2)^{-1}]$$

where Pco₂ was calculated from the standard gas' known CO₂ concentration.

Hemolymph analysis

Tco₂ was measured using a total CO₂ analyzer (Capnicon 5, Cameron Instruments). The pH was measured using a blood gas meter (BGM200, Cameron Instruments) with pH glass and reference electrodes (E301, E351, Cameron Instruments). The pH electrodes were installed in a water jacket and maintained at experiment temperatures. Hemolymph Pco₂ and [HCO₃⁻] were calculated by rearranging the Henderson-Hasselbalch equation^{19,21)}. The *aco*₂ and pK_{app} obtained here were then used for hemolymph Pco₂ calculation

from pH and T_{CO_2} :

$$P_{CO_2} = T_{CO_2} \cdot [a_{CO_2} \cdot (1 + 10^{(pH - pK_{app})})]^{-1}$$

$[HCO_3^-]$ was calculated from T_{CO_2} , a_{CO_2} , and P_{CO_2} , or from a_{CO_2} , P_{CO_2} , pH, and pK_{app} using the equation:

$$[HCO_3^-] = T_{CO_2} - a_{CO_2} \cdot P_{CO_2}$$

$$[HCO_3^-] = a_{CO_2} \cdot P_{CO_2} \cdot 10^{(pH - pK_{app})}$$

For assessment of the relationship between hemolymph pH and $[HCO_3^-]$ in the experimental animals, the non-bicarbonate buffer values (β_{NB}) were calculated from the slope of the relational expression between pH and $[HCO_3^-]$.

Statistical analysis

All data are expressed as mean \pm standard error. The Kruskal-Wallis test was performed for changes in hemolymph sample properties and the calculated a_{CO_2} or pK_{app} . Multiple comparison for all pairs used the Steel-Dwass test. Statistically significant differences were set at $P < 0.05$. All analyses were carried out using the statistical software Kyplot v. 6.0 (KyensLab Inc., Japan).

Results

The mean hemolymph a_{CO_2} values between 10°C and 20°C are 44.4 – 60.1 $\mu\text{M/L/torr}$ (Table 1) and are significantly different ($P < 0.05$). A polynomial equation was fitted to the a_{CO_2} data obtained at the experimental temperatures (Fig. 1). The equation of a_{CO_2} and temperature

is expressed as follows:

$$a_{CO_2} = 74.005 - 1.2936 \cdot T - 0.00944 \cdot T^2$$

where T is the temperature, and the parameter units in the equations are $\mu\text{M/L/torr}$ for a_{CO_2} and °C for T.

The hemolymph pK_{app} at known P_{CO_2} (standard gases), with corresponding measured pH and T_{CO_2} are shown for each temperature in Table 2-4. The changes in pH, T_{CO_2} , and pK_{app} between 10°C and 20°C significantly associated with the increase in P_{CO_2} ($P < 0.05$), except for pK_{app} at 10°C. The distribution of pK_{app} mean values and the corresponding pH are shown for each temperature in Fig. 2. The mean values of pK_{app} at 20°C were higher than that at 10°C and 15°C ($P < 0.05$). The mean hemolymph pK_{app} values were 5.99896 ± 0.01936 at 10°C, 6.06434 ± 0.02193 at 15°C, and 6.32955 ± 0.03880 at 20°C. The polynomial equation was fitted to the mean pK_{app} values at the experimental temperatures (Fig. 3). The relationship between pK_{app} and temperature is expressed as follows:

$$pK_{app} = 6.4675 - 0.08682 \cdot T + 0.003996 \cdot T^2$$

where T is temperature in °C.

The hemolymph pH and calculated $[HCO_3^-]$ values are listed in Table 5. The non-bicarbonate buffer values (β_{NB}), obtained as a regression coefficient relating pH and $[HCO_3^-]$, were 2.5 Slykes at 10°C, 2.2 Slykes at 15°C, and 3.4 Slykes at 20°C.

Table 1. Disk abalone *Haliotis (Nordotis) discus discus* hemolymph CO_2 solubility (a_{CO_2}) at three water temperatures

WT (°C)	CO_2 solubility ($\mu\text{M/L/torr}$)		
	Mean	SE	N
10	60.1	3.08	6
15	52.5	1.59	6
20	44.4	0.79	5

Table 2. Mean values of measured pH, total CO₂ content (Tco₂) and calculated apparent dissociation constant of carbonic acid (pKapp) of disk abalone *Haliotis (Nordotis) discus discus* hemolymph with known Pco₂ standard gases at 10°C

Standard gas		Hemolymph			
CO ₂	Pco ₂	pH	Tco ₂	pKapp	N
(%)	(torr)		(mM/L)		
0.102	0.770	7.396	1.20	6.0061130	5
0.203	1.54	7.237	1.40	6.0946147	5
1.01	7.65	6.751	3.26	5.9680550	5
2.00	15.1	6.526	4.00	5.9902950	5
5.01	37.8	6.182	6.41	5.9270075	5

Barometric pressure, 766.6 ± 6.6 torr; water temperature, 10.3 ± 0.3°C (Mean ± SD).
Mean value of pKapp, 5.99896 ± 0.019368.

Table 3. Mean values of measured pH, total CO₂ content (Tco₂) and calculated apparent dissociation constant of carbonic acid (pKapp) of disk abalone *Haliotis (Nordotis) discus discus* hemolymph with known Pco₂ standard gases at 15°C

Standard gas		Hemolymph			
CO ₂	Pco ₂	pH	Tco ₂	pKapp	N
(%)	(torr)		(mM/L)		
0.203	1.53	7.320	1.388	6.125411	6
0.515	3.87	7.050	1.860	6.151023	6
1.01	7.60	6.834	2.736	6.067520	6
2.00	15.0	6.576	3.657	6.018804	6
5.01	37.6	6.220	5.586	5.958987	6

Barometric pressure, 765 ± 3.3 torr; water temperature, 15.0 ± 0.1°C (Mean ± SD).
Mean value of pKapp, 6.06434 ± 0.021930.

Table 4. Mean values of measured pH, total CO₂ content (Tco₂) and calculated apparent dissociation constant of carbonic acid (pKapp) of disk abalone *Haliotis (Nordotis) discus discus* hemolymph with known Pco₂ standard gases at 20°C

Standard gas		Hemolymph			
CO ₂	Pco ₂	pH	Tco ₂	pKapp	N
(%)	(torr)		(mM/L)		
0.203	1.51	7.500	1.043	6.258071	6
0.515	3.83	7.361	1.808	6.292781	6
1.01	7.51	7.232	2.623	6.307798	6
2.00	14.9	7.144	3.472	6.418255	6
5.01	37.2	6.803	4.877	6.397612	6

Barometric pressure, 760 ± 1.6 torr; water temperature, 20.0 ± 0.2°C (Mean ± SD).
Mean value of pKapp, 6.32955 ± 0.03880.

Table 5. Mean values of measured pH and calculated bicarbonate concentration ($[HCO_3^-]$) of disk abalone *Haliotis (Nordotis) discus discus* hemolymph with known P_{CO_2} standard gases at 10-20°C

Standard gas	10°C		15°C		20°C	
	pH	$[HCO_3^-]$ (mM/L)	pH	$[HCO_3^-]$ (mM/L)	pH	$[HCO_3^-]$ (mM/L)
CO ₂ (%)						
0.102	7.396	1.15	—	—	—	—
0.203	7.237	1.30	7.320	1.31	7.471	1.56
0.515	—	—	7.050	1.66	7.232	2.02
1.01	6.751	2.78	6.834	2.34	7.295	2.89
2.00	6.526	3.13	6.576	2.87	7.092	3.15
5.01	6.182	4.078	6.220	3.61	6.913	3.51

The non-bicarbonate buffer value (β_{NB}), 2.5 Slykes at 10°C; 2.2 Slykes at 15°C; 3.4 Slykes at 20°C.

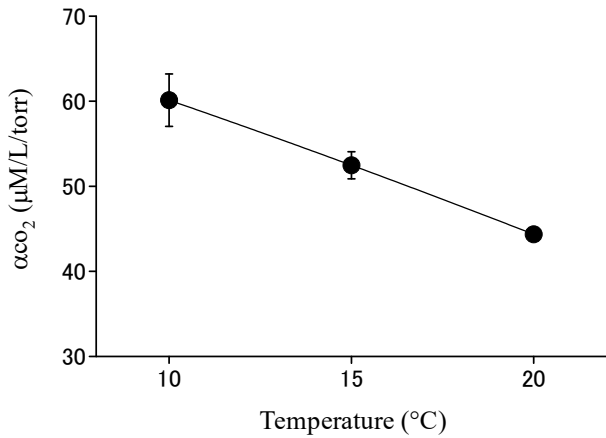


Fig. 1 Influence of temperature on CO_2 solubility coefficient (α_{CO_2}) for disk abalone *Haliotis (Nordotis) discus discus* hemolymph between 10°C and 20°C. Data are Mean \pm SE. Solid lines are fitted to the data and the equation: $\alpha_{CO_2} = 74.005 - 1.2936 \cdot T - 0.00944 \cdot T^2$ ($R^2 = 0.9997$).

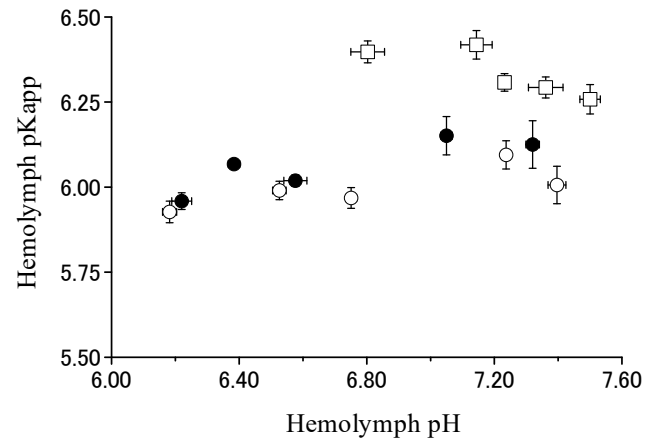


Fig. 2 Mean value distributions for pK_{app} and corresponding pH of the hemolymph in disk abalone *Haliotis (Nordotis) discus discus* at each temperature. Data are Mean \pm SE. Open circle: 10°C; solid circle: 15°C; open square: 20°C. The mean values of pK_{app} at 20°C were higher than that at 10°C and 15°C ($P < 0.05$, Steel-Dwass multiple comparison test).

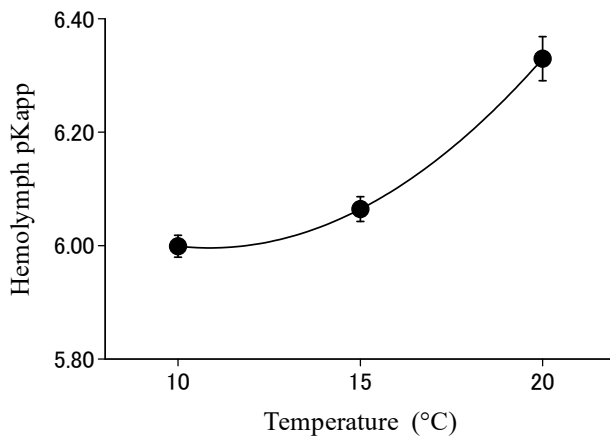


Fig. 3 Influence of temperature on the apparent carbonic acid dissociation constant (pK_{app}) of disk abalone *Haliotis (Nordotis) discus discus* hemolymph between 10°C and 20°C. Data are Mean \pm SE. The curved line is fitted to the data and the equation: $pK_{app} = 6.4675 - 0.08686 \cdot T + 0.003996 \cdot T^2$ ($R^2 = 0.8915$).

Discussion

We undertook a preliminary investigation of the effect of temperature on disk abalone hemolymph a_{CO_2} and pKapp, and clarified these relationships between 10°C and 20°C. Disk abalone hemolymph a_{CO_2} was 44–60 $\mu\text{M}/\text{L}/\text{torr}$ at the experimental temperature. Cameron (1986) reported CO_2 solubility as a function of temperature and salinity, with solubility coefficients of 44.74–60.80 $\mu\text{M}/\text{L}/\text{torr}$ at a salinity of 30 (psu) between 10°C and 20°C²². The obtained hemolymph a_{CO_2} of the disk abalone between 10°C and 20°C reflects the coefficients reported by Cameron (1986). With increasing temperature, the hemolymph a_{CO_2} decreased, with significantly different values at each temperature evaluated. Fitted to the disk abalone hemolymph a_{CO_2} data at each experimental temperature we obtained the polynomial equation (Fig. 1). Thus, the hemolymph a_{CO_2} equation enables a_{CO_2} estimation in the temperature range between 10°C and 20°C.

The distributions of the hemolymph pKapp differed with the corresponding pH at each temperature (Fig. 2). The pKapp at 20°C was higher than that at other temperatures, with no significant differences in pKapp distribution between 10°C and 15°C. The mean pKapp value was 5.999 at 10°C, 6.064 at 15°C, and 6.330 at 20°C. The pKapp value is equal to the pH at which it is most effective as a buffer²³. Thus, the disk abalone hemolymph increases the buffer effectiveness at higher temperatures. This phenomenon may relate to seasonal variations in chemical properties of the hemolymph, such as ion or proteins. Although there are few reports of disk abalone hemolymph pKapp values, other reported marine bivalve hemolymph pKapp values include 6.114 for *Mytilus edulis* at 12°C^{24,25}, 6.073 for *Crassostrea gigas* at 23°C²⁶, 6.064 for *Mimamclamys nobilis* at 24°C²⁷, 5.998 for *Pinctada margaritifera* at 26°C²⁸, and 5.819 for *P. fucata martensii* at 28°C²⁹. As the hemolymph pKapp value observed in this study was higher than those reported for bivalves, the disk abalone hemolymph appears a more effective buffer compared with the bivalves. The polynomial equation (Fig. 3) was obtained by fitting the disk abalone

hemolymph pKapp data obtained at various experimental temperatures. Thus, hemolymph pKapp may be estimated for temperatures between 10°C and 20°C.

The hemolymph $[\text{HCO}_3^-]$ in disk abalone was calculated using the hemolymph a_{CO_2} and pKapp. The regression coefficient relating $[\text{HCO}_3^-]$ and the pH was the non-bicarbonate buffer value. The β_{NB} at 20°C was 3.4 Slykes, and higher than that at 10–15°C (2.2–2.5 Slykes). The non-bicarbonate buffer value was determined by the buffer capacity of the non-bicarbonate buffer system (for example, protein buffer system), and used to quantify the amount of buffering of the solution component^{23,30}. In disk abalone, hemolymph pH changes at 20°C are predicted to require greater quantities of acid or base compared to the lower temperatures. Thus, at 20°C, the disk abalone may be better able to maintain hemolymph pH.

According to the public data of Japan Meteorological Agency, the coastal sea surface temperatures from Ibaragi prefecture to Kyusyu, Nagasaki prefecture are 13–28°C³¹, which is the habitat of the disk abalone. Komazawa et al. (2004a, 2004b) compared disk abalone juvenile growth rates in different water temperatures (13–28°C), reporting the highest juvenile growth rate was at 13°C and 16–19°C^{3,4}. Yamamoto et al. (2011) reported the effect of temperature on adult disk abalone respiration, suggesting an upper limit of suitable water temperature between 22°C and 25 °C from gill ventilation volume and oxygen uptake measurements¹⁵. In this study, the hemolymph acid–base balance was examined between 10°C and 20°C. This temperature range was narrower than the conditions of the previous reports and will be expanded to evaluate the acid–base equilibrium at other temperatures. Boutilier et al. (1985) described a_{CO_2} and pKapp variations with temperature and ionic strength¹⁹. As hemolymph pKapp changes were influenced by the pH, it is necessary to study the relationships among temperature, pH, and pKapp of the disk abalone hemolymph.

Acknowledgments

We would like to express our sincere gratitude to Dr. Ken-ichi Yamamoto, Professor Emeritus, for securing the experimental animals for this study.

References

- 1) Okutani T: Haliotidae. *In*: Okutani T (ed) Marine Mollusks in Japan (2nd edition), Tokai University Press, Kanagawa, 773-774 (2017)
- 2) Ishida O: Effect of population density on the growth of juveniles of the abalone, *Haliotis discus discus*. *Suisanzoshoku (Aquaculture Science)*, **41**, 431-433 (1993)
- 3) Komazawa I, Kudo M, Sugino T: Growth of three abalone species reared under different water temperatures of 22, 24, 26, and 28°C. *Report of the Tokyo Metropolitan Fisheries Experiment Station*, **213**, 57-65 (2004a)
- 4) Komazawa I, Kudo M, Sugino T: Growth of two abalone species reared under different water temperatures of 13, 16, 19, and 22°C. *Report of the Tokyo Metropolitan Fisheries Experiment Station*, **213**, 67-75 (2004b)
- 5) Segawa S: Preliminary experiment on the effect of temperature on rates of oxygen consumption and ammonia excretion of young disk abalone, *Nordotis discus discus*. *Suisanzoshoku (Aquaculture Science)*, **43**, 219-224 (1995)
- 6) Nakatsugawa T: Infectious nature of a disease in cultured juvenile abalone with muscular atrophy. *Fish Pathology*, **25**, 207-211 (1990)
- 7) Nakatsugawa T, Okabe M, Muroga K: Horizontal transmission of amyotrophia in Japanese black abalone. *Fish Pathology*, **35**, 11-14 (2000)
- 8) Okada K, Nishimura M, and Kawamura T: Prevention of amyotrophia in mass production of 0-year-old abalone, *Haliotis discus*, by Quarantine. *Suisanzoshoku (Aquaculture Science)*, **48**, 657-663 (2000)
- 9) Shibata T, Chikushi Y, Nakamoto T, Watanabe K, Nagashima T: Prevention of amyotrophia in large scale production of juvenile abalone, *Haliotis discus discus*, by ultraviolet irradiation of water supply. *Suisanzoshoku (Aquaculture Science)*, **50**, 227-232 (2002)
- 10) Zoysa DM, Whang I, Lee Y, Lee S, Lee JS, Lee J: Defensin from disk abalone *Haliotis discus discus*: Molecular cloning, sequence characterization and immune response against bacterial infection. *Fish & Shellfish Immunology*, **28**, 261-266 (2010)
- 11) Bathige SDNK, Umasuthan N, Godahewa GI, Thulasitha WS, Whang I, Won SH, Kim C, Lee J: Two variants of selenium-dependent glutathione peroxidase from the disk abalone *Haliotis discus discus*: Molecular characterization and immune responses to bacterial and viral stresses. *Fish & Shellfish Immunology*, **45**, 648-655 (2015)
- 12) Yamamoto K, Handa T, Kondo M: Structure of the digestive diverticula of abalone *Haliotis (Nordotis) discus discus*. *Journal of National Fisheries University*, **53**, 105-116 (2005)
- 13) Yamamoto K, Handa T, Kondo M: Ctenidium structure of the abalone *Haliotis (Nordotis) discus discus* (Gastropoda: Aspidobranchia). *Journal of National Fisheries University*, **56**, 287-298 (2008)
- 14) Yamamoto K, Handa T: New method of direct measurement of ventilation volume of abalones (Mollusca, Haliotidae). *Journal of National Fisheries University*, **49**, 59-65 (2001)
- 15) Yamamoto K, Handa T, Tsunoji T: Effect of seasonal rise in water temperature on the respiration in the abalone *Haliotis (Nordotis) discus discus*. *Aquaculture Science*, **59**, 529-534 (2011)
- 16) Yamamoto K, Handa T: Flow through respiratory pores of the abalone *Haliotis (Nordotis) discus discus*. *Aquaculture Science*, **60**, 393-396 (2012)
- 17) Yamamoto K, Handa T: Effect of hypoxia on oxygen uptake in the abalone *Haliotis (Nordotis) discus discus*. *Journal of National Fisheries University*, **61**, 81-85 (2013)
- 18) Handa T, Araki A, Yamamoto K: Acid-base balance of hemolymph in disk abalone *Haliotis (Nordotis) discus discus* in normoxia conditions. *Journal of National Fisheries University*, **67**, 133-139 (2019).
- 19) Boutilier RG, Iwama GK, Heming TA, Randall DJ: The apparent pK of carbonic acid in rainbow trout blood plasma between 5 and 15 °C. *Respiratory Physiology*, **61**, 237-254 (1985)

- 20) Namba K, Kobayashi M, Aida S, Uematsu K, Yoshida M, Kondo Y, Miyata U: Persistent relaxation of the adductor muscle of oyster *Crassostrea gigas* induced by magnesium ion. *Fisheries Science*, **61**, 241-244 (1995)
- 21) Davenport HW: Fundamental Equation. *In*: Davenport HW (ed) *The ABC of Acid-base Chemistry* (6th edition), University of Chicago Press, Chicago, 39-41 (1974)
- 22) Cameron JN: The Solubility of Carbon Dioxide as a Function of Temperature and Salinity (Appendix Table). *In*: Cameron JN (ed) *Principles of Physiological Measurement*, Academic Press, London, 258-259 (1986)
- 23) Heisler N: Acid-base Regulation, Interrelationships between Gaseous and Ionic Exchange. *In*: Boutilier RG (ed) *Vertebrate Gas Exchange, Comparative & Environmental physiology 6*, Springer-Verlag Berlin & Heidelberg, 211-251 (1990)
- 24) Booth CE, McDonald DG, Walsh PJ: Acid-base balance in the sea mussel, *Mytilus edulis*. I. Effects of hypoxia and air-exposure on hemolymph acid-base status. *Marine Biology Letters*, **5**, 347-358 (1984)
- 25) Lindinger MI, Lauren DJ, McDonald DG: Acid-base balance in the sea mussel, *Mytilus edulis*. III. Effects of environmental hypercapnia on intra- and extracellular acid-base balance. *Marine Biology Letters*, **5**, 371-381 (1984)
- 26) Handa T, Araki A, Yamamoto K: Acid-base balance of the hemolymph in Pacific oyster *Crassostrea gigas* in normoxic conditions. *Journal of National Fisheries University*, **66**, 103-110 (2018)
- 27) Handa T, Yamamoto K: Estimation of CO₂ partial pressure and bicarbonate concentration in the hemolymph of the noble scallop *Mimachlamys nobilis*. *Journal of National Fisheries University*, **64**, 188-194 (2016)
- 28) Handa T, Yamamoto K: Estimation of CO₂ partial pressure and bicarbonate concentration in the hemolymph of the black-lip pearl oyster *Pinctada margaritifera*. *Journal of National Fisheries University*, **63**, 181-188 (2015)
- 29) Handa T, Yamamoto K: The acid-base balance of the hemolymph in the pearl oyster *Pinctada fucata martensii* under normoxic conditions. *Aquaculture Science*, **60**, 113-117 (2012)
- 30) Claiborne JB: Acid-base Regulation. *In*: Evans DH (ed) *The Physiology of Fishes* (2nd edition), CRC Press LLC, Florida, 177-198 (1998)
- 31) Japan Meteorological Agency: <https://www.data.jma.go.jp/kaiyou/data/db/kaikyo/series/engan/engan.html> (2023)